

## ***Brilliant camouflage: photonic crystals in the diamond weevil, *Entimus imperialis****

Bodo D. Wilts, Kristel Michielsen, Jeroen Kuipers, Hans De Raedt and Doekele G. Stavenga

*Proc. R. Soc. B* 2012 **279**, 2524-2530 first published online 29 February 2012

doi: 10.1098/rsjb.2011.2651

---

### **Supplementary data**

["Data Supplement"](#)

<http://rsjb.royalsocietypublishing.org/content/suppl/2012/02/23/rsjb.2011.2651.DC1.html>

### **References**

[This article cites 36 articles, 20 of which can be accessed free](#)

<http://rsjb.royalsocietypublishing.org/content/279/1738/2524.full.html#ref-list-1>

### **Email alerting service**

Receive free email alerts when new articles cite this article - sign up in the box at the top right-hand corner of the article or click [here](#)

# Brilliant camouflage: photonic crystals in the diamond weevil, *Entimus imperialis*

Bodo D. Wilts<sup>1,\*</sup>, Kristel Michielsen<sup>2</sup>, Jeroen Kuipers<sup>3</sup>,

Hans De Raedt<sup>1</sup> and Doekele G. Stavenga<sup>1</sup>

<sup>1</sup>Computational Physics, Zernike Institute for Advanced Materials, University of Groningen, Groningen 9747AG, The Netherlands

<sup>2</sup>Institute for Advanced Simulation, Jülich Supercomputing Centre, Research Centre Jülich, Jülich 52425, Germany

<sup>3</sup>Department of Cell Biology, University Medical Centre Groningen, University of Groningen, Groningen 9713AV, The Netherlands

The neotropical diamond weevil, *Entimus imperialis*, is marked by rows of brilliant spots on the overall black elytra. The spots are concave pits with intricate patterns of structural-coloured scales, consisting of large domains of three-dimensional photonic crystals that have a diamond-type structure. Reflectance spectra measured from individual scale domains perfectly match model spectra, calculated with anatomical data and finite-difference time-domain methods. The reflections of single domains are extremely directional (observed with a point source less than 5°), but the special arrangement of the scales in the concave pits significantly broadens the angular distribution of the reflections. The resulting virtually angle-independent green coloration of the weevil closely approximates the colour of a foliaceous background. While the close-distance colourful shininess of *E. imperialis* may facilitate intersexual recognition, the diffuse green reflectance of the elytra when seen at long-distance provides cryptic camouflage.

**Keywords:** structural colour; photonic bandgap materials; scatterometry; diffuse reflection; communication

## 1. INTRODUCTION

Animal coloration is due to spectrally selective light reflections on the outer body parts [1,2]. The resulting coloration mostly serves a biological function in intra- and interspecific signalling [3,4], thus improving mating chances [5], but it can also be used for camouflage in the animal's native habitat [6–8]. Generally, two types of coloration are recognized, pigmentary and structural. Pigmentary (or chemical) coloration occurs when pigments absorb incoherently scattered light in a restricted wavelength range. Structural (or physical) coloration is due to nanometre-sized structures with periodically changing refractive indices, causing coherent light scattering. Pigmentary coloration is by far the most common in the animal kingdom, but structural coloration is widely encountered as well, and not seldom structural colours are modified by spectrally filtering pigments [1,2,9].

If the structures causing the physical colours are regular with a periodicity in the order of the wavelength of visible light, the materials are referred to as photonic crystals [10]. One-dimensional photonic crystals consist of parallel thin film layers of alternating high and low refractive index materials, i.e. the well-known multi-layers. They create the metallic and polarized reflections of, for example, the skin of cephalopods [9] and fishes [11], the elytra of jewel beetles [12–15], scarabs [16,17], and the breast feathers of birds of paradise [18]. Two-dimensional

photonic crystals, that is, structures with periodicity in two dimensions, underlie the coloration of peacock feathers [19,20]. Three-dimensional photonic crystals have been found in the scales of many weevils and beetles [1,21–24], but also in butterflies [25–28]. Quasi-ordered three-dimensional photonic crystal structures, which are periodic in all three dimensions, although imperfect, have been identified in bird feathers [29] and in the scales of some coleopterans [30].

The colorations of the elytra of weevils and beetles are especially diverse [1,2]. Here, we investigate the diamond weevil, *Entimus imperialis*, which is endemic to the neotropical regions, especially southwest Brazil [31]. *Entimus imperialis* is a member of the monophyletic genus *Entimus* (Curculinoidea: Entiminae: Entimini). The relatively large weevils of this genus, with a 15–45 mm body length, have strikingly iridescent scales, immersed in pits on the weevil's elytra and legs (figure 1a). A unique property of the elytral scales of *E. imperialis* is the presence of extremely large, crystalline domains, which allows an in-depth analysis of their physical properties. In a previous study, we identified the domains to be single-network diamond-type photonic crystals [22]. Here, we present reflectance spectra of single domains together with calculated reflectance spectra of the domains, based on their anatomical dimensions. We find that perfect matches of the experimental and computationally derived spectra can be obtained for appropriate orientations of the crystalline structure. We thus demonstrate, for the first time to our knowledge, that the optical properties of the weevil scales can be quantitatively understood.

Curiously, the scales are arranged inside concave pits. This leads to a drastic change in appearance of the animal

\* Author for correspondence ([b.d.wilts@rug.nl](mailto:b.d.wilts@rug.nl)).

Electronic supplementary material is available at <http://dx.doi.org/10.1098/rspb.2011.2651> or via <http://rsob.royalsocietypublishing.org>.

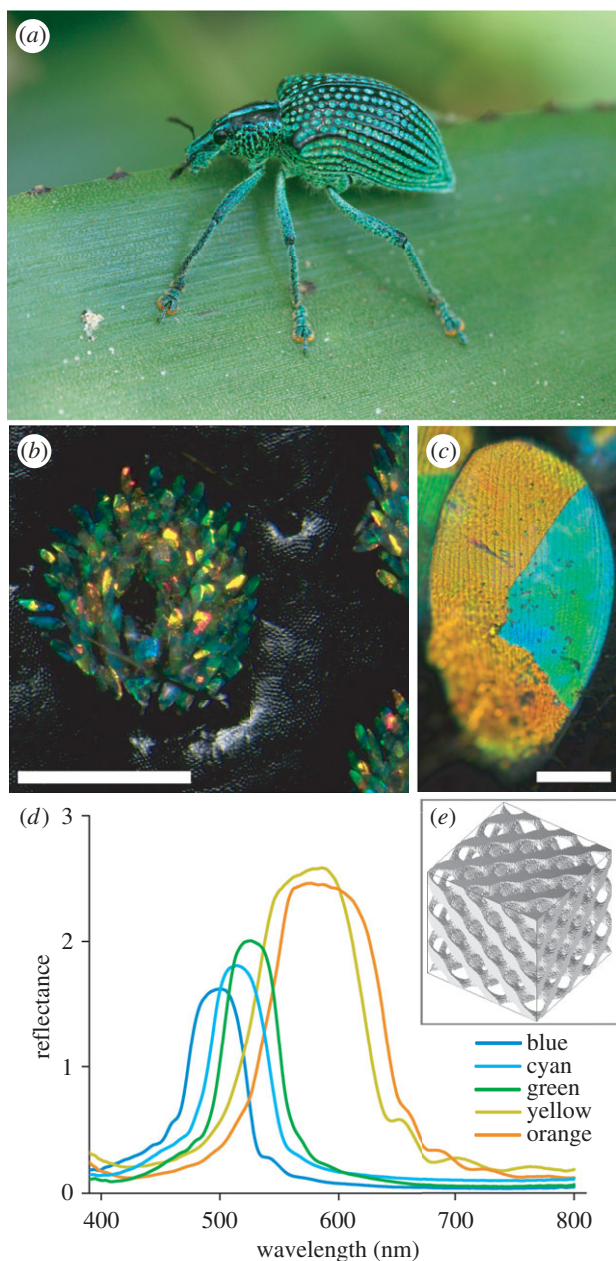


Figure 1. The diamond weevil, *Entimus imperialis*. (a) The intact animal with its black elytra studded with numerous yellow-green pits, sitting on a green leaf (image courtesy of Carlos M. Ribeiro). (b) A single pit as seen in an epi-illumination microscope (bar: 0.5 mm). (c) A single scale with a few differently coloured domains (bar: 20 μm). (d) Reflectance spectra of single, differently coloured domains measured with a microspectrophotometer, applying white-light illumination at about normal incidence, with a white diffuser as a reference. (e) Rendered model of the single-diamond photonic crystal found inside the scale lumen.

when observed at long distance or from close-up. Measurement of the integral reflectance of the scale assembly in the elytral pits yielded spectra matching the spectrum of a green, foliaceous background, the weevil's natural habitat [32]. This suggests that the scale set is optimized for camouflage for distant predators. However, the concentration of the glittering scales in distinct pits causes a spotted patterning for observers at close range. The bright patterning may allow ready recognition for nearby conspecifics.

## 2. MATERIAL AND METHODS

### (a) Animals

A specimen of the diamond weevil, *E. imperialis*, of the Coleoptera collection in the Natural History Museum Naturalis (Leiden, The Netherlands; curator J. Krikken), was photographed with a Canon EOS-30D camera. Extensive investigations were performed on a specimen obtained from Prof. J.-P. Vigneron (University of Namur, Belgium).

### (b) Spectrophotometry

Reflectance spectra of single domains were measured with a microspectrophotometer connected to an AvaSpec-2048-2 spectrometer (Avantes, Eerbeek, The Netherlands); spectra of intact wings were acquired with an integrating sphere (Avantes Avasphere-50-Refl) connected to the Avantes spectrometer. The light source was a xenon or a deuterium-halogen (Avantes D(H)-S) lamp. For all reflectance measurements, a white diffuse reflectance tile (Avantes WS-2) served as a reference.

### (c) Imaging scatterometry

We examined the far-field, 180° hemispherical angular distribution of the light scattered by single scales as well as by a complete pit. We, therefore, used an imaging scatterometer (ISM), which is built around an ellipsoidal mirror [33,34].

### (d) Anatomy

The photonic structure inside the wing scales was investigated by transmission electron microscopy (TEM), using a FEI CM 100 transmission electron microscope. The wing scales were embedded in a mixture of Epon and Araldite following a standard embedding procedure [12].

### (e) Modelling

The light scattering by single-network diamond photonic crystals was simulated for a number of differently oriented crystals with three-dimensional finite-difference time-domain (FDTD) calculations. We used TDME3D, a massively parallel Maxwell equation solver [6]. We modelled the chitin-air diamond network by the refractive index  $n(X,Y,Z) = 1.55 + 0.06i$  [35,36] for the volume filled with chitin (filling function  $f(X,Y,Z) < -0.5$ ) and  $n(X,Y,Z) = 1$  for the volume filled with air ( $f(X,Y,Z) \geq -0.5$ ). For a single-diamond lattice, the filling function  $f(X,Y,Z)$  is given by  $f(X,Y,Z) = \cos Z \sin(X+Y) + \sin Z \cos(X-Y)$  [25], where  $X = 2\pi x/a$ ,  $Y = 2\pi y/a$  and  $Z = 2\pi z/a$ , with  $x$ ,  $y$  and  $z$  denoting the coordinates in the cubic structure. We took for the lattice constant  $a = 445$  nm. The simulation boxes had a linear dimension of  $16a$  and the modelled diamond structure thickness of  $6a$ , closely matching the experimentally observed thickness. We used perfectly matched-layer boundaries for the simulation box. To mimic the experimental orientations, cross sections of single scales, given by Miller indices  $(hkl)$  of the single-diamond structure, were oriented in the simulation box perpendicular to the direction of light incidence. One simulation run, which is the calculation for one structural orientation, one wavelength, one polarization and one incidence angle, required a memory of approx. 60 GB. The simulations were performed on the IBM BlueGene/P of the University of Groningen.

## 3. RESULTS

### (a) Optical appearance of the diamond weevil scales

The diamond weevil, *E. imperialis*, has overall black elytra, which are marked by rows of yellow-green

glittering spots (figure 1*a,b*). With only minor changes of the viewing angle, the shiny spots feature strikingly different colours, which give them a diamond-like appearance. The strong directionality of the reflections indicates that the origin of these local, vivid colours is structural [2,3].

A closer look at the elytral pits shows that the diamond-like spots are assemblies of highly directionally reflective scales covering the walls of concave pits in the elytra (figure 1*b*). Similar brilliant scales are also found on the thorax and the legs of the weevil (figure 1*a*; see also [31]). Single wing scales have an elongated shape, with length 100  $\mu\text{m}$  and width 50  $\mu\text{m}$ . The scales consist of coloured domains, with colour ranging from turquoise to yellow-orange. Most scales feature a few (three to five) large domains (figure 1*c*); a minority of scales have only one domain.

The coloured domains, when observed with an epillumination microscope at high magnification, resolve into striped patterns with orientations depending on the colour of the domains (figure 1*c*; see also fig. 2*a* of Wilts *et al.* [22]). The striped, grating-like patterns with different orientations strongly suggested that the scale lumen contains differently ordered crystalline structures. In a separate study, we identified the structures inside the scales by using a novel optical characterization technique, namely hemispherical Brillouin zone imaging [22]. We found that the scale interior consists of an ordered, three-dimensional lattice of chitin, enveloped by an external cuticular cortex. The air-chitin assembly forms a three-dimensional single-network diamond photonic crystal with face-centred cubic lattice symmetry [22]. The scales have a chitin-filling fraction of approximately 0.3 and a lattice constant  $445 \pm 10$  nm (figure 1*e*).

Biological photonic crystals like that of the *Entimus* weevil have a low refractive index contrast (for air-chitin assemblies: 1.56) [35,36]. Consequently, the spatial direction and spectral distribution of the light reflected by the crystal depends on the orientation of the crystal with respect to the direction of illumination [6,10]. In other words, the different-coloured domains of the weevil's scales are due to differently oriented photonic crystals (figure 1*b,c*) [22].

### (b) Reflectance spectra measured from single-scale domains

To quantify the observed scale colours, we measured the reflectance spectra of single-scale domains with a microspectrophotometer. This yielded a set of representative narrow-band reflectance spectra with half-width (full width at half maximum (FWHM)) approximately 50–80 nm and peak wavelengths at approximately 480 nm (turquoise-blue), approximately 520 nm (cyan), approximately 540 nm (green) and between 560 and 610 nm (yellow-orange). The amplitude of the measured spectra increased with the peak wavelength (figure 1*d*).

### (c) Orientations of single domains and modelling of the photonic response

For a quantitative understanding of the photonic response of a single domain, its crystal orientation inside the scale has to be known. We, therefore, performed TEM on single scales. Figure 2 (left column) shows the most commonly encountered crystal orientations in single-scale

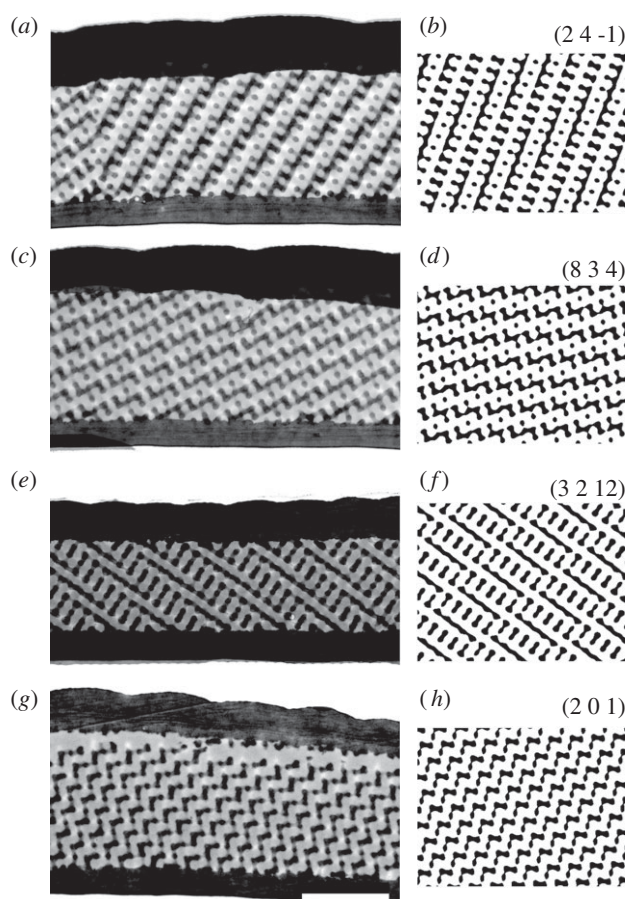


Figure 2. TEM of single scales revealing different orientations. The left column (*a,c,e,g*) shows a side-view of four different scales (bar: 2  $\mu\text{m}$ , in *g*). The right column (*b,d,f,h*) shows the matching crystal orientations of a single-diamond crystal with a chitin-filling fraction approx. 0.3 and thickness  $0.2a$  ( $a$  is the lattice constant). The orientations are given by the Miller indices ( $h k l$ ) above each simulated cross section.

cross sections. We compared the TEM images, obtained from approximately 70 nm thick sections, with computer-generated, projections modelled by level surfaces equivalent to 70 nm thick sections of single-diamond photonic crystal structures. We, therefore, rotated the modelled sections stepwise until the computationally created projection matched the pattern of the experimentally obtained TEM sections (figure 2, right column). Matching images could only be obtained for a single-diamond crystal, not for gyroid or simple primitive crystals [27]. Fast Fourier transforms of the TEM images and the matching computer-generated images yielded the lattice constant of the diamond crystal:  $a = 445 \pm 10$  nm as well as the Miller indices of the orientation of the crystal. Miller indices ( $h k l$ ) give the directions and planes of equal symmetry of a crystal with respect to the standard unit cell, i.e. the normal crystal orientation with respect to the scale surface [10].

Having derived the lattice constant,  $a = 445$  nm, we subsequently calculated reflectance spectra for the differently-oriented, single-diamond photonic crystals. We used a parallel Maxwell solver and calculated the reflectance for TE-(transverse electric, or s-) as well as for TM-(transverse magnetic, or p-) polarized light (figure 3; electronic supplementary material, S1). The simulated reflectance spectra had a half-width (FWHM) approximately

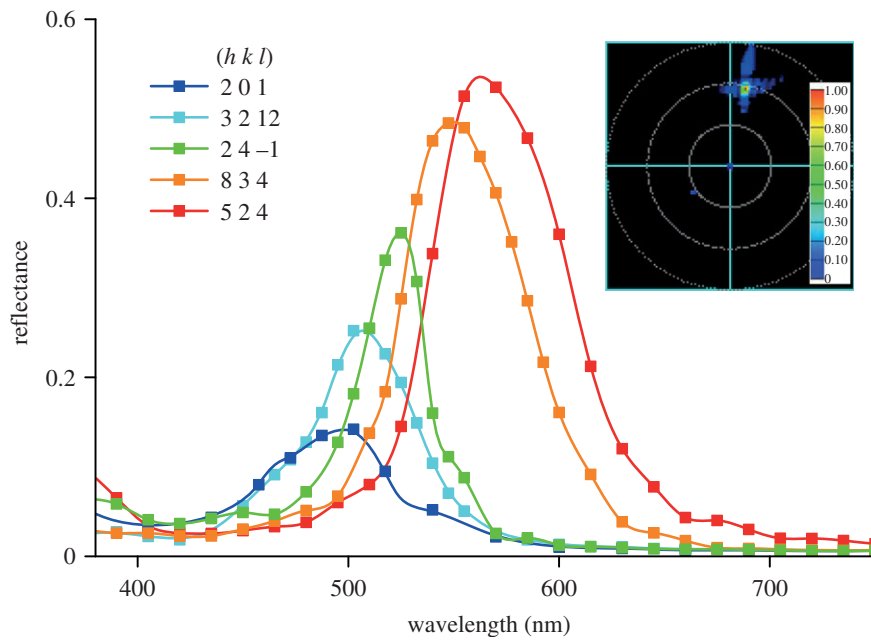


Figure 3. Reflectance spectra calculated with FDTD for a diamond-type photonic crystal with orientation denoted by the Miller indices  $(hkl)$  when exposed to unpolarized light. For this, spectra calculated for TE- and TM-polarized light were averaged (electronic supplementary material, figure S1). The lattice constant of the single diamond-type crystal was set to  $a = 445$  nm. The inset shows the calculated scattering diagram for the (8 3 4)-domain for normal-incident, TM-polarized light with wavelength 550 nm. The scattered light is concentrated in a very narrow spatial angle (the circles indicate scattering angles of  $30^\circ$ ,  $60^\circ$  and  $90^\circ$ , respectively).

30–80 nm and peaked in the blue to yellow-orange wavelength range, from 490 to 580 nm. For each orientation, given by the Miller indices in figure 3 and electronic supplementary material, S1, the TE- and TM-reflectance spectra differed only very slightly in peak wavelength (approx. 20 nm) and peak amplitude (approx. 0.1). We, therefore, averaged the TE- and TM-reflectance spectra calculated for the various orientations of the crystal domains (figure 3). The experimental reflectance spectra of single domains, which were measured with unpolarized light (figure 1*d*), closely correspond to the averaged calculated spectra.

#### (d) Effect of the scale arrangement on spatial visibility

The FDTD calculations predicted very directional, mirror-like reflections for the individual crystal orientations (see inset of figure 3). To determine the spatial reflection properties of the scales, and especially to assess how the scale arrangement in elytral pits affects the spatial scale reflection properties, we performed ISM. For this, we glued both single scales and small pieces of elytra with scale-carrying pits to the tip of pulled micropipettes and illuminated them with a narrow aperture, white light beam with a variable spot size.

Figure 4 presents the scatterograms resulting from differently sized illumination areas; a single domain (spot-size diameter  $d = 15$   $\mu\text{m}$ ; figure 4*a*), a single scale ( $d = 50$   $\mu\text{m}$ ; figure 4*b*), a few scales ( $d = 140$   $\mu\text{m}$ ; figure 4*c*) and the complete pit ( $d = 800$   $\mu\text{m}$ ; figure 4*d*). Illumination of single domains (figure 4*a*) resulted in light-scattering profiles with a distinct colour and a very narrow solid angle. The spatial extent of the reflected spots, half-width of  $5$ – $10^\circ$ , was virtually identical to the aperture of the illumination

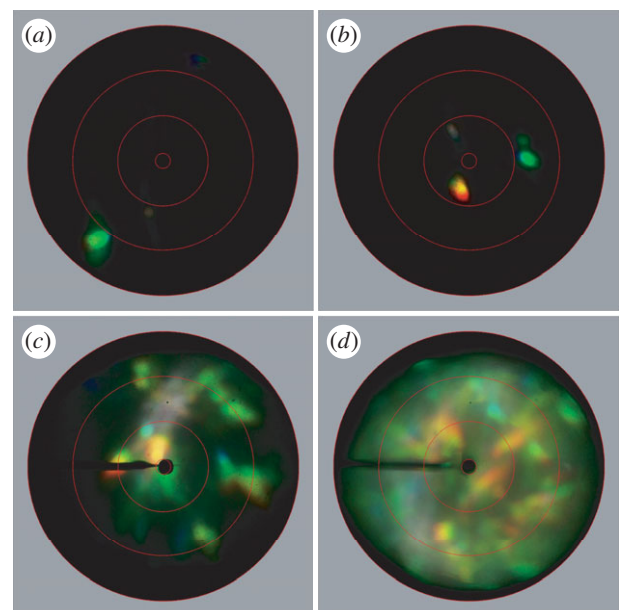


Figure 4. Imaging scatterometry of single domains, single scales and multiple scales illuminated with a narrow-aperture light beam (approx.  $5^\circ$ ). The illumination spot size was (a) 15  $\mu\text{m}$ , (b) 50  $\mu\text{m}$ , (c) 140  $\mu\text{m}$  and (d) 800  $\mu\text{m}$ . (a) A single domain creates a scatter pattern with a very narrow-aperture of approximately  $5^\circ$ . (b) Illumination of a single scale with two domains leads to two narrow reflection spots. (c) Illumination of four to five scales leads to a multitude of spots, or to a spatially extended scatter pattern, determined by the number of domains in the scale. (d) Illumination of all scales in one pit results in a scatter pattern that covers the whole hemisphere, with a number of distinct spots over a more or less uniform green-yellow background. The red circles indicate scattering angles of  $5^\circ$ ,  $30^\circ$ ,  $60^\circ$  and  $90^\circ$  respectively.

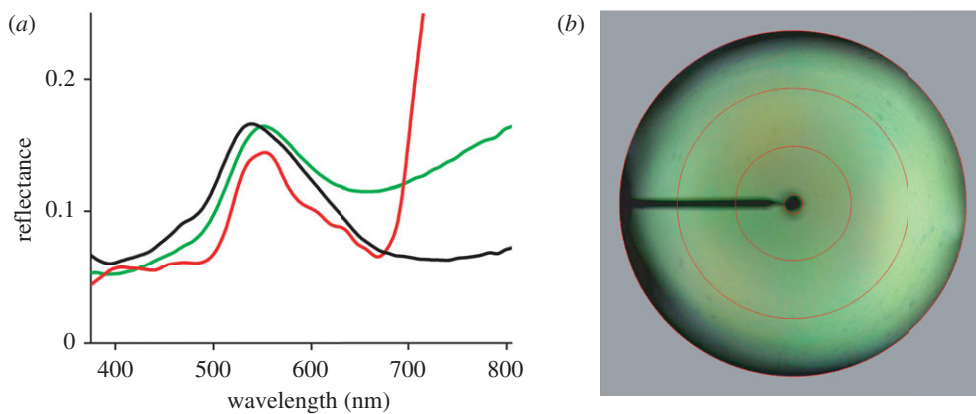


Figure 5. Brilliant camouflage. (a) Reflectance spectra measured with an integrating sphere of an elytron of the diamond weevil, *Entimus imperialis*, of a green oak leaf, and of the wing underside of the green hairstreak butterfly, *Callophrys rubi*. (b) Scatter pattern of a single-scale-carrying pit in a weevil elytron illuminated with wide-aperture white light. Black solid line, *E. imperialis*; green line, *C. rubi*; red line, green leaf. The red circles indicate scattering angles of 5°, 30°, 60° and 90° respectively.

beam, which was approximately 5° [33]. Strikingly, the angular position of the reflected spot severely differed between the domains, meaning that the directions of the reflected light beams severely deviated from each other (figure 4a; see also inset figure 3b for the simulated reflectance pattern of a single domain).

Increasing the spot size increased the complexity of the reflection pattern. Illumination of only a restricted number of scales, each having a small number of different domains, caused a scatter pattern that was the sum of the directional reflections of the single domains (figure 4b,c). An increase in the size of the illuminated area increased the number of reflected spots in the scatterogram. When a whole elytral pit was illuminated, further broadening of the reflection pattern occurred (figure 4d). The scatterograms then contained a few prominent reflection spots on a more or less angle-independent green-yellow background.

#### (e) Large-area reflections of the weevil elytra

Under normal circumstances, light will be scattered by numerous elytral scales, not by single scales or single domains. The reflectance spectrum measured from a large area will therefore be the weighted sum of the spectra of the illuminated individual domains with different orientations (figure 4). To assess the overall colour reflectance of the diamond weevil, we measured the reflectance spectrum of the elytra with an integrating sphere, which had a large illumination spot size, approximately 8 mm, thus capturing approximately 10 elytral pits. As expected, the resulting reflectance spectrum, peak wavelength approximately 540 nm and half-width approximately 180 nm, was considerably broader than the spectra of single domains (figure 5a). Interestingly, the reflectance spectrum of the large illumination spot strongly resembles the reflectance spectra of green leaves. Wide-angle illumination of a single pit also results in an approximately angle-independent green coloration (figure 5b).

## 4. DISCUSSION

### (a) Biophotonic structures in weevils and beetles

The elytral scales of the weevil *E. imperialis* have uniquely large domains, which allow spectral measurements on individual domains. Both the directionality of the reflected

light and the measured reflectance spectra are in full agreement with predictions based on the derived anatomical data and the structure of diamond-type photonic crystals.

Photonic crystals with a diamond structure seem to be restricted to beetles and weevils (Coleoptera) [1], whereas photonic crystals with a gyroid structure have so far only been encountered in lycaenid and papilionid butterflies [6,25,27]. In nascent butterfly wing scales, specific interactions of the cell plasma membrane and the intracellular smooth endoplasmic reticulum initiate the development of single gyroid networks by cubic membrane folding via a double gyroid intermediate [27,37]. To date, relatively little is known about the development of nascent weevil wing scales. Concerning the diamond structures, we may speculate that the developmental pathway involves a double-diamond intermediate, as this intermediate is observed as a stable phase in cubic membrane folding [38,39]. The presence of diamond-type photonic crystals in weevil scales may have distinct evolutionary advantages that can range from differences in reflectance properties to enhancement of mating chances [5]. Light reflected by gyroid photonic crystals can be highly polarized [28], whereas diamond photonic crystals show hardly any polarization dependency (figure 3; [10,22,26]). Whether or not, this difference in polarization properties has a biological function has yet to be investigated.

### (b) Biological implications of the scale arrangement

Single-scale domains exhibit strong reflections in narrow spatial angles (figure 4a,b). The reflecting domains in the scale assembly in an elytral pit together create a distinct, vividly coloured spot. The distance between the elytral pits is approximately 1 mm, so that, assuming a spatial resolution of the weevil eyes of approximately 1° [40], potential mates can discriminate adjacent spots from a distance as far as approximately 6 cm. The near-field brilliancy of the dotted pattern of the weevil's elytra may therefore be used as a mate recognition signal [3].

The scales in a pit all have a similar orientation. Their tips are directed towards the elytral apex, corresponding to the general scale patterning of weevil elytra. However, the scales at the slopes of the conical pits have different angular orientations and this causes a spatially broadened reflection signal. The pit will therefore act effectively as a signal scrambler, by creating an overall multi-domain

arrangement comparable to that observed in single scales of the weevil *L. augustus* [23] or the ventral wing scales of the green hairstreak butterfly, *Callophrys rubi* [6,27]; in these cases, the domain size is smaller by a factor of about ten.

Overall, the array of coloured pits gives rise to a uniform, diffusely green-yellow appearance when illuminated with a wide-angled light source (figure 5*b*). The integral yellow-green reflectance closely mimics the reflectance of green leaves (generally leaf spectra resemble the spectrum of an oak leaf, shown as an example in figure 5*b*). This immediately suggests that the coloured scales in the concave pits serve to camouflage the weevil against a mainly green background (see also figure 1*a*), especially for their common predators, birds [40]. Further, the dotted arrangement of the pits on the elytra will support camouflage by disruptive patterning, a common mechanism to achieve camouflage [8], e.g. applied by cuttlefish [7,41].

The coloured scales of *E. imperialis* are concentrated in concave pits on its black elytra. Clearly, the scales are thus not vulnerable to mechanical wear. The concentration of the scales in pits, therefore, presumably serve three functions: the scales are protected from mechanical damage and accidental loss, the pattern of brilliant spots facilitates intraspecific recognition, and the integral green coloration favours camouflage in foliaceous environments [3,4].

We thank H. L. Leertouwer and A. J. M. Vey for collaboration and P. Vukusic for reading an early version of this manuscript. This study was financially supported by AFOSR/EOARD (grant FA8655-08-1-3012) and NCF, The Netherlands.

## REFERENCES

- Seago, A. E., Brady, P., Vigneron, J. P. & Schultz, T. D. 2009 Gold bugs and beyond: a review of iridescence and structural colour mechanisms in beetles (Coleoptera). *J. R. Soc. Interface* **6**(Suppl 2), S165–S184. (doi:10.1098/rsif.2008.0354.focus)
- Kinoshita, S. 2008 *Structural colors in the realm of nature*. Singapore: World Scientific.
- Doucet, S. M. & Meadows, M. G. 2009 Iridescence: a functional perspective. *J. R. Soc. Interface* **6**(Suppl 2), S115–S132. (doi:10.1098/rsif.2008.0395.focus)
- Meadows, M. G., Butler, M. W., Morehouse, N. I., Taylor, L. A., Toomey, M. B., McGraw, K. J. & Rutowski, R. L. 2009 Iridescence: views from many angles. *J. R. Soc. Interface* **6**(Suppl 2), S107–S113. (doi:10.1098/rsif.2009.0013.focus)
- Darwin, C. 1859 *The origin of species by means of natural selection, or the preservation of favoured races in the struggle for life*. London, UK: John Murray.
- Michielsen, K., De Raedt, H. & Stavenga, D. G. 2010 Reflectivity of the gyroid biophotonic crystals in the ventral wing scales of the green hairstreak butterfly, *Callophrys rubi*. *J. R. Soc. Interface* **7**, 765–771. (doi:10.1098/rsif.2009.0352)
- Buresch, K. C., Mäthger, L. M., Allen, J. J., Bennice, C., Smith, N., Schram, J., Chiao, C. C., Chubb, C. & Hanlon, R. T. 2011 The use of background matching versus masquerade for camouflage in cuttlefish *Sepia officinalis*. *Vis. Res.* **51**, 2362–2368. (doi:10.1016/j.visres.2011.09.009)
- Stevens, M. & Merilaita, S. 2009 Defining disruptive coloration and distinguishing its functions. *Phil. Trans. R. Soc. B* **364**, 481–488. (doi:10.1098/rstb.2008.0216)
- Mäthger, L. M. & Hanlon, R. T. 2007 Malleable skin coloration in cephalopods: selective reflectance, transmission and absorbance of light by chromatophores and iridophores. *Cell Tissue Res.* **329**, 179–186. (doi:10.1007/s00441-007-0384-8)
- Joannopoulos, J. D. 2008 *Photonic crystals: molding the flow of light*, 2nd edn. Princeton, NJ: Princeton University Press.
- Mäthger, L. M., Land, M. F., Siebeck, U. E. & Marshall, N. J. 2003 Rapid colour changes in multilayer reflecting stripes in the paradise whiptail, *Pentapodus paradiseus*. *J. Exp. Biol.* **206**, 3607–3613. (doi:10.1242/jeb.00599)
- Stavenga, D. G., Wilts, B. D., Leertouwer, H. L. & Hariyama, T. 2011 Polarized iridescence of the multilayered elytra of the Japanese jewel beetle, *Chrysochroa fulgidissima*. *Phil. Trans. R. Soc. B* **366**, 709–723. (doi:10.1098/rstb.2010.0197)
- Hariyama, T., Hironaka, M., Takaku, Y., Horiguchi, H. & Stavenga, D. G. 2005 The leaf beetle, the jewel beetle, and the damselfly; insects with a multilayered show case. In *Structural color in biological systems: principles and applications* (eds S. Kinoshita & S. Yoshioka), pp. 153–176. Osaka, Japan: Osaka University Press.
- Mason, C. W. 1927 Structural colors in insects. III. *J. Phys. Chem.* **31**, 1856–1872. (doi:10.1021/j150282a008)
- Durrer, H. & Villiger, W. 1972 Schillerfarben von *Euchroma gigantea* (L.): (Coleoptera: Buprestidae): Elektronenmikroskopische Untersuchung der Elytra. *Int. J. Insect Morphol. Embryol.* **1**, 233–240. (doi:10.1016/0020-7322(72)90031-1)
- Sharma, V., Crne, M., Park, J. O. & Srinivasarao, M. 2009 Structural origin of circularly polarized iridescence in jeweled beetles. *Science* **325**, 449–451. (doi:10.1126/science.1172051)
- Brady, P. & Cummings, M. 2010 Differential response to circularly polarized light by the jewel scarab beetle *Chrysinia gloriosa*. *Am. Nat.* **175**, 614–620. (doi:10.1086/651593)
- Stavenga, D. G., Leertouwer, H. L., Marshall, N. J. & Osorio, D. 2011 Dramatic colour changes in a bird of paradise caused by uniquely structured breast feather barbules. *Proc. R. Soc. B* **278**, 2098–2104. (doi:10.1098/rspb.2010.2293)
- Zi, J., Yu, X., Li, Y., Hu, X., Xu, C., Wang, X., Liu, X. & Fu, R. 2003 Coloration strategies in peacock feathers. *Proc. Natl Acad. Sci. USA* **100**, 12 576–12 578. (doi:10.1073/pnas.2133313100)
- Loyau, A., Gomez, D., Moureau, B., Thery, M., Hart, N. S., Saint Jalme, M., Bennett, A. T. D. & Sorci, G. 2007 Iridescent structurally based coloration of eyespots correlates with mating success in the peacock. *Behav. Ecol.* **18**, 1123–1131. (doi:10.1093/beheco/arm088)
- Parker, A. R., Welch, V. L., Driver, D. & Martini, N. 2003 Structural colour: opal analogue discovered in a weevil. *Nature* **426**, 786–787. (doi:10.1038/426786a)
- Wilts, B. D., Michielsen, K., De Raedt, H. & Stavenga, D. G. In press. Hemispherical Brillouin zone imaging of a diamond-type biological photonic crystal. *J. R. Soc. Interface*. (doi:10.1098/rsif.2011.0730)
- Galusha, J. W., Richey, L. R., Gardner, J. S., Cha, J. N. & Bartl, M. H. 2008 Discovery of a diamond-based photonic crystal structure in beetle scales. *Phys. Rev. E* **77**, 050 904. (doi:10.1103/PhysRevE.77.050904)
- Welch, V., Lousse, V., Deparis, O., Parker, A. & Vigneron, J. P. 2007 Orange reflection from a three-dimensional photonic crystal in the scales of the weevil *Pachyrhynchus congestus pavonius* (Curculionidae). *Phys. Rev. E* **75**, 041919. (doi:10.1103/PhysRevE.75.041919)

- 25 Michielsen, K. & Stavenga, D. G. 2008 Gyroid cuticular structures in butterfly wing scales: biological photonic crystals. *J. R. Soc. Interface* **5**, 85–94. (doi:10.1098/rsif.2007.1065)
- 26 Poladian, L., Wickham, S., Lee, K. & Large, M. C. 2009 Iridescence from photonic crystals and its suppression in butterfly scales. *J. R. Soc. Interface* **6**(Suppl 2), S233–S242. (doi:10.1098/rsif.2008.0353.focus)
- 27 Saranathan, V., Osuji, C. O., Mochrie, S. G., Noh, H., Narayanan, S., Sandy, A., Dufresne, E. R. & Prum, R. O. 2010 Structure, function, and self-assembly of single network gyroid ( $I4_132$ ) photonic crystals in butterfly wing scales. *Proc. Natl Acad. Sci. USA* **107**, 11 676–11 681. (doi:10.1073/pnas.0909616107)
- 28 Saba, M., Thiel, M., Turner, M. D., Hyde, S. T., Gu, M., Grosse-Brauckmann, K., Neshev, D. N., Mecke, K. & Schröder-Turk, G. E. 2011 Circular dichroism in biological photonic crystals and cubic chiral nets. *Phys. Rev. Lett.* **106**, 103 902. (doi:10.1103/PhysRevLett.106.103902)
- 29 Shawkey, M. D., Saranathan, V., Palsdottir, H., Crum, J., Ellisman, M. H., Auer, M. & Prum, R. O. 2009 Electron tomography, three-dimensional Fourier analysis and colour prediction of a three-dimensional amorphous biophotonic nanostructure. *J. R. Soc. Interface* **6**, S213–S220. (doi:10.1098/rsif.2008.0374.focus)
- 30 Dong, B. Q., Liu, X. H., Zhan, T. R., Jiang, L. P., Yin, H. W., Liu, F. & Zi, J. 2010 Structural coloration and photonic pseudogap in natural random close-packing photonic structures. *Opt. Express* **18**, 14 430–14 438. (doi:10.1364/OE.18.014430)
- 31 Morrone, J. J. 2002 The Neotropical weevil genus *Entimus* (Coleoptera: Curculionidae: Entiminae): cladistics, biogeography, and modes of speciation. *Coleopt. Bull.* **56**, 501–513. (doi:10.1649/0010-065X(2002)056[0501:TNWGEC]2.0.CO;2)
- 32 Mc Kay, F., Oleiro, M., Cabrera Walsh, G., Gandolfo, D., Cuda, J. P. & Wheeler, G. S. 2009 Natural enemies of Brazilian peppertree (Sapindales: Anacardiaceae) from Argentina: their possible use for biological control in the USA. *Fla. Entomol.* **92**, 292–303. (doi:10.1653/024.092.0213)
- 33 Stavenga, D. G., Leertouwer, H. L., Piri, P. & Wehling, M. F. 2009 Imaging scatterometry of butterfly wing scales. *Opt. Exp.* **17**, 193–202. (doi:10.1364/OE.17.000193)
- 34 Vukusic, P. & Stavenga, D. G. 2009 Physical methods for investigating structural colours in biological systems. *J. R. Soc. Interface* **6**(Suppl 2), S133–S148. (doi:10.1098/rsif.2008.0386.focus)
- 35 Vukusic, P., Sambles, J. R., Lawrence, C. R. & Wootton, R. J. 1999 Quantified interference and diffraction in single *Morpho* butterfly scales. *Proc. R. Soc. B* **266**, 1403–1411. (doi:10.1098/rspb.1999.0794)
- 36 Leertouwer, H. L., Wilts, B. D. & Stavenga, D. G. 2011 Refractive index and dispersion of butterfly chitin and bird keratin measured by polarizing interference microscopy. *Opt. Exp.* **19**, 24 061–24 066. (doi:10.1364/OE.19.024061)
- 37 Ghiradella, H. 1989 Structure and development of iridescent butterfly scales: lattices and laminae. *J. Morphol.* **202**, 69–88. (doi:10.1002/jmor.1052020106)
- 38 Hyde, S., Andersson, S., Larsson, K., Blum, Z., Landh, T., Lidin, S. & Ninham, B. 1997 *The language of shape: the role of curvature in condensed matter: physics, chemistry, and biology*. Amsterdam, The Netherlands: Elsevier.
- 39 Almsherqi, Z. A., Kohlwein, S. D. & Deng, Y. 2006 Cubic membranes: a legend beyond the flatland of cell membrane organization. *J. Cell Biol.* **173**, 839–844. (doi:10.1083/jcb.200603055)
- 40 Osorio, D. & Vorobyev, M. 2005 Photoreceptor spectral sensitivities in terrestrial animals: adaptations for luminance and colour vision. *Proc. R. Soc. B* **272**, 1745–1752. (doi:10.1098/rspb.2005.3156)
- 41 Cuthill, I. C., Stevens, M., Sheppard, J., Maddocks, T., Parraga, C. A. & Troscianko, T. S. 2005 Disruptive coloration and background pattern matching. *Nature* **434**, 72–74. (doi:10.1038/nature03312)